

Production of Antimicrobial Peptides in *Nicotiana benthamiana* Plants for Efficient Treatments

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Antibiotic-resistant infections are projected to cause over 39 million global deaths between 2025 and 2050, with treatment costs exceeding \$66 billion annually. Rather than replacing ineffective antibiotics, it is possible to restore them. Plant-produced antimicrobial peptides (AMPs) offer a promising yet untapped synergistic treatment. However, the absence of critical pre-clinical evaluations has hindered AMP's clinical translation. This study aims to express AMPs in *Nicotiana benthamiana* using a tobacco mosaic virus (TMV) vector for evaluation of purity, antibacterial, cytotoxicity, and synergistic properties. A TMV-AMP plasmid was inserted into agrobacteria for agroinfiltration into *N. benthamiana*. After TMV-AMP plant infection, AMPs were purified via RP-HPLC. AMP viability was assessed via PCR, an antibacterial assay on *E. coli* MG1655, a cytotoxicity assay on human embryonic kidney (HEK) and human dermal fibroblast (HDF) cells, and a synergy assay combining AMPs with azithromycin against resistant *E. coli* PI-7. AMPs exhibited antibacterial activity against *E. coli* MG1655, with a minimum inhibitory concentration of 50 µg/mL, achieving $92.46\% \pm 4.53$ bacterial inhibition. AMPs showed minimal cytotoxicity to HEK and HDF, maintaining >90% cell viability up to 50 µg/mL. The optimal AMP concentration for minimal cytotoxicity and maximal antibacterial activity was 50 µg/mL, with a final plant-produced yield of 2.1 mg/mL, the highest reported for plant-TMV expression. AMPs enhanced azithromycin's efficacy against *E. coli* PI-7; a combination costing less than 1 cent per dose. This study highlights the potential of clinically translating AMPs as a cost-effective and clinically viable treatment to restore antibiotic efficacy, potentially saving lives by 2050.

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